



## Journal club

Omidreza Firuzi, MD, PhD
Professor of Pharmacology
Medicinal and natural products chemistry research center
19-09-1399

#### Resource



## A Landscape of Pharmacogenomic Interactions in Cancer

Francesco Iorio,1,2,20 Theo A. Knijnenburg,3,4,20 Daniel J. Vis,4,20 Graham R. Bignell,2,20 Michael P. Menden,1,5,20 Michael Schubert,1 Nanne Aben,4,6 Emanuel Gonçalves,1 Syd Barthorpe,2 Howard Lightfoot,2 Thomas Cokelaer,1,2,17 Patricia Greninger,7 Ewald van Dyk,4 Han Chang,8 Heshani de Silva,8 Holger Heyn,9 Xianming Deng,10,11,18 Regina K. Egan,7 Qingsong Liu,10,11 Tatiana Mironenko,2 Xeni Mitropoulos,7 Laura Richardson,2 Jinhua Wang,10,11 Tinghu Zhang,10,11 Sebastian Moran,9 Sergi Sayols,9,19 Maryam Soleimani,2 David Tamborero,12 Nuria Lopez-Bigas,12,13 Petra Ross-Macdonald,8 Manel Esteller,9,13,14 Nathanael S. Gray,10,11 Daniel A. Haber,7,15 Michael R. Stratton,2 Cyril H. Benes,7 Lodewyk F.A. Wessels,4,6,16,21 Julio Saez-Rodriguez,1,5,21 Ultan McDermott,2,21,\* and Mathew J. Garnett2,21,\*

<sup>1</sup>European Molecular Biology Laboratory, European Bioinformatics Institute, Wellcome Genome Campus, Cambridge CB10 1SA, UK <sup>2</sup>Wellcome Trust Sanger Institute, Wellcome Genome Campus, Cambridge CB10 1SA, UK

<sup>&</sup>lt;sup>3</sup>Institute for Systems Biology, Seattle, WA 98109, USA

Division of Molecular Carcinogenesis, The Netherlands Cancer Institute, Amsterdam 1066 CX, The Netherlands

<sup>&</sup>lt;sup>5</sup>Faculty of Medicine, Joint Research Centre for Computational Biomedicine, RWTH Aachen University, Aachen 52057, Germany

<sup>&</sup>lt;sup>6</sup>Department of EEMCS, Delft University of Technology, Delft 2628 CD, the Netherlands

<sup>&</sup>lt;sup>7</sup>Center for Cancer Research, Massachusetts General Hospital, Harvard Medical School, Charlestown, MA 02129, USA

<sup>&</sup>lt;sup>8</sup>Genetically Defined Diseases and Genomics, Bristol-Myers Squibb Research and Development, Hopewell, NJ 08534, USA

Gancer Epigenetics and Biology Program (PEBC), Bellvitge Biomedical Research Institute (IDIBELL), L'Hospitalet 08908, Barcelona, Catalonia, Spain

<sup>&</sup>lt;sup>10</sup>Department of Cancer Biology, Dana-Farber Cancer Institute, Boston, MA 02215, USA

<sup>&</sup>lt;sup>11</sup>Department of Biological Chemistry & Molecular Pharmacology, Harvard Medical School, Boston, MA 02215, USA

<sup>&</sup>lt;sup>12</sup>Research Program on Biomedical Informatics, IMIM Hospital del Mar Medical Research Institute and Universitat Pompeu Fabra, Barcelona 08003, Spain

<sup>&</sup>lt;sup>13</sup>Institució Catalana de Recerca i Estudis Avançats (ICREA), 08010 Barcelona, Catalonia, Spain

<sup>&</sup>lt;sup>14</sup>Department of Physiological Sciences II of the School of Medicine, University of Barcelona, L'Hospitalet 08908, Barcelona, Catalonia, Spain

<sup>&</sup>lt;sup>15</sup>Howard Hughes Medical Institute, Chevy Chase, MD 20815, USA

<sup>&</sup>lt;sup>16</sup>Cancer Genomics Netherlands, Uppsalalaan 8, Utrecht 3584CT, the Netherlands

<sup>&</sup>lt;sup>17</sup>Present address: Bioinformatics and Biostatistics Hub, C3BI, USR 3756 IP CNRS, Institut Pasteur, 75015 Paris, France

<sup>&</sup>lt;sup>18</sup>Present address: Innovation Center for Cell Signaling Network, School of Life Sciences, Xiamen University, 361102 Xiamen, China

<sup>&</sup>lt;sup>19</sup>Present address: Institute of Molecular Biology, Mainz 55128, Germany

<sup>20</sup>Co-first author

<sup>21</sup> Co-senior author

<sup>\*</sup>Correspondence: um1@sanger.ac.uk (U.M.), mg12@sanger.ac.uk (M.J.G.) http://dx.doi.org/10.1016/j.cell.2016.06.017

## ARTICLE

## Systematic identification of genomic markers of drug sensitivity in cancer cells

Mathew J. Garnett<sup>1</sup>\*, Elena J. Edelman<sup>2</sup>\*, Sonja J. Heidorn<sup>1</sup>\*, Chris D. Greenman<sup>1</sup>†, Anahita Dastur<sup>2</sup>, King Wai Lau<sup>1</sup>, Patricia Greninger<sup>2</sup>, I. Richard Thompson<sup>1</sup>, Xi Luo<sup>2</sup>, Jorge Soares<sup>1</sup>, Qingsong Liu<sup>3,4</sup>, Francesco Iorio<sup>1,5</sup>, Didier Surdez<sup>6</sup>, Li Chen<sup>2</sup>, Randy J. Milano<sup>2</sup>, Graham R. Bignell<sup>1</sup>, Ah T. Tam<sup>2</sup>, Helen Davies<sup>1</sup>, Jesse A. Stevenson<sup>2</sup>, Syd Barthorpe<sup>1</sup>, Stephen R. Lutz<sup>2</sup>, Fiona Kogera<sup>1</sup>, Karl Lawrence<sup>1</sup>, Anne McLaren-Douglas<sup>1</sup>, Xeni Mitropoulos<sup>2</sup>, Tatiana Mironenko<sup>1</sup>, Helen Thi<sup>2</sup>, Laura Richardson<sup>1</sup>, Wenjun Zhou<sup>3,4</sup>, Frances Jewitt<sup>1</sup>, Tinghu Zhang<sup>3,4</sup>, Patrick O'Brien<sup>1</sup>, Jessica L. Boisvert<sup>2</sup>, Stacey Price<sup>1</sup>, Wooyoung Hur<sup>3,4</sup>, Wanjuan Yang<sup>1</sup>, Xianming Deng<sup>3,4</sup>, Adam Butler<sup>1</sup>, Hwan Geun Choi<sup>3,4</sup>, Jae Won Chang<sup>3,4</sup>, Jose Baselga<sup>2</sup>, Ivan Stamenkovic<sup>7</sup>, Jeffrey A. Engelman<sup>2</sup>, Sreenath V. Sharma<sup>2</sup>†, Olivier Delattre<sup>6</sup>, Julio Saez-Rodriguez<sup>5</sup>, Nathanael S. Gray<sup>3,4</sup>, Jeffrey Settleman<sup>2</sup>, P. Andrew Futreal<sup>1</sup>, Daniel A. Haber<sup>2,8</sup>, Michael R. Stratton<sup>1</sup>, Sridhar Ramaswamy<sup>2</sup>, Ultan McDermott<sup>1</sup> & Cyril H. Benes<sup>2</sup>

<sup>1</sup>Cancer Genome Project, Wellcome Trust Sanger Institute, Hinxton CB10 1SA, UK. <sup>2</sup>Massachusetts General Hospital Cancer Center, Harvard Medical School, Charlestown, Massachusetts 02129, USA. <sup>3</sup>Department of Cancer Biology, Dana Farber Cancer Institute, 44 Binney Street, Boston Massachusetts 02115, USA. <sup>4</sup>Department of Biological Chemistry and Molecular Pharmacology, Harvard Medical School, 250 Longwood Avenue, Boston, Massachusetts 02115, USA. <sup>5</sup>EMBL-EBI, Wellcome Trust Genome Campus, Cambridge CB10 1SD, UK. <sup>6</sup>Laboratoire de génétique et biologie des cancers, Institut Curie, 75248 Paris, Cedex 05, France. <sup>7</sup>Division of Experimental Pathology, Institute of Pathology, Centre Hospitalier Universitaire Vaudois (CHUV), 1005 Lausanne, Switzerland. <sup>8</sup>Howard Hughes Medical Institute, Chevy Chase, Maryland 20815, USA. †Present addresses: Department of Computing, University of East Anglia, Norwich NR4 7TJ, UK (C.D.G.); The Genome Analysis Centre, Norwich Research Park, Norwich NR4 7UH, UK (C.D.G.); Oncology Drug Discovery, Novartis Institutes for Biomedical Research, 250 Massachusetts Avenue, Cambridge, Massachusetts 02139, USA (S.V.S.).

\*These authors contributed equally to this work.



SCIENCE

PEOPLE

INNOVATION

**NEWS** 

Group leads Core team Previous team members Associated research Related groups Programmes and Facilities Partners More V

**ABOUT** 

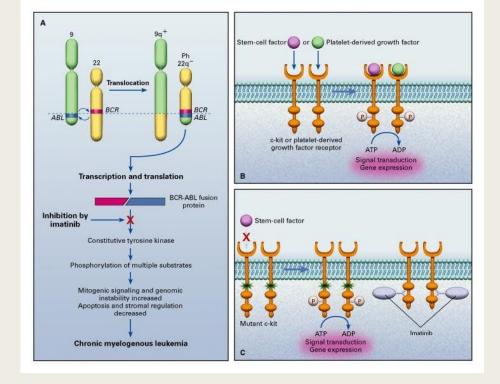


## THE CANCER GENOME ATLAS

• Studies from The Cancer Genome Atlas (TCGA) and the International Cancer Genome Consortium (ICGC) have generated comprehensive catalogs of the cancer genes involved in tumorigenesis across a broad range of cancer types (Lawrence et al., 2014; Tamborero et al., 2013b; Zack et al., 2013).

## Successful examples

- Imatinib selectively targets the protein product of the BCR-ABL translocation in chronic myeloid leukaemia (CML) has revolutionized the treatment of CML.
- However, a large number of cancer drugs have not been linked to specific genomic alterations.
- Systematic methods to identify **predictive biomarkers** during their early development could have a profound effect on the success of new cancer drug development.



## EGFR example: low frequency determinants of clinical response

- Clinical responses to small-molecule selective inhibitors of the epidermal growth factor receptor (EGFR), erlotinib and gefitinib were seen in a small fraction of treated patients with non-small-cell lung cancer (NSCLC)
- These are patients with mutations within the EGFR kinase domain (10% to 20% of patients)
- This shows the importance of **stratifying patients** with NSCLC for EGFR TKI therapy based on the mutational status of *EGFR* in their tumors.
- It is also clear that *EGFR* activating mutations **are not the sole determinants of clinical response**.

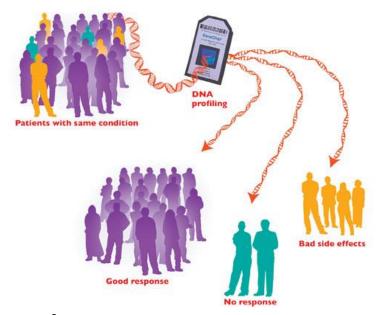
Table 1. Oncogenic Kinases As Therapeutic Targets in Cancer						
Kinase	Alteration	Tumor Types	Therapeutic Agent	Study		
Receptor tyrosine kinases				24		
EGFR	Mutation, amplification	Lung, GBM	Gefitinib, erlotinib	Lynch et al <sup>24</sup>		
ErbB2/HER2	Amplification	Breast	Lapatinib	Gomez et al <sup>20</sup>		
FGFR1	Translocation	CML	PKC412, BIBF 1120	Hilberg et al <sup>22</sup>		
FGFR2	Amplification, mutation	Gastric, breast, endometrial	PKC412, BIBF 1120	Hilberg et al <sup>22</sup>		
FGFR3	Translocation, mutation	Multiple myeloma	PKC412, BIBF 1120	Hilberg et al <sup>22</sup>		
PDGFR $\alpha$	Mutation	GBM, GIST	Sunitinib, sorafenib, imatinib	Cohen et al, <sup>17</sup> Demetri et al <sup>18</sup>		
PDGFR $eta$	Translocation	CMML	Sunitinib, sorafenib, imatinib	Cohen et al, <sup>17</sup> Demetri et al <sup>18</sup>		
ALK	Mutation/amplification	Lung, neuroblastoma, ALCL	PF-2341066	Christensen et al <sup>16</sup>		
MET	Amplification	Gefitinib-resistant NSCLC, gastric	PF-2341066, XL184, SU11274	Salgia et al, <sup>26</sup> Zou et al <sup>30</sup>		
IGF-1R	Activation by IGF-II ligand	Colorectal, pancreatic	CP 751 871, AMG479	Hewish et al <sup>21</sup>		
c-KIT	Mutation	GIST	Sunitinib, imatinib	Demetri et al <sup>18</sup>		
FLT3	Internal tandem duplication	AML	Lestaurtinib, XL999	Illmer and Ehninger <sup>23</sup>		
RET	Mutation, translocation	Thyroid medullary carcinoma	XL184	Salgia et al <sup>26</sup>		
Nonreceptor tyrosine kinases						
Abl	Translocation (Bcr-Abl)	CML	Imatinib	Druker et al <sup>19</sup>		
JAK2	Mutation (V617F), translocation	CML, MPD	Lestaurtinib, INCB018424	Verstovsek et al <sup>28</sup>		
Src	Overexpression	NSCLC, ovarian, breast, sarcoma	KX2-391, dasatinib, AZD0530	Blume-Jensen et al <sup>14</sup>		
Serine/threonine/lipid kinases						
BRAF	Mutation (V600E)	Melanoma, colon	SB-590885, PLX-4720, RAF265, XL281	Smalley and Flaherty <sup>27</sup>		
Aurora kinase A and B	Overexpression	Breast, colon, leukemia	MK-5108 (VX-689)	Warner et al <sup>29</sup>		
Polo-like kinases	Overexpression	Breast, lung, lymphoma, colon	BI2536, GSK461364	Warner et al <sup>29</sup>		
mTOR	Increased activation	Renal cell carcinoma	Temsirolimus (CCI-779), BEZ235	Chan et al, <sup>15</sup> Maira et al <sup>25</sup>		
PI3K	PIK3CA mutations	Colorectal, breast, GBM, gastric	BEZ235	Maira et al <sup>25</sup>		

### McDermott et al 2019 JCO

### Predictive biomarkers

- Systematic studies of cancer genomes have provided unprecedented insights into the molecular nature of cancer.
- Clinical responses to anticancer therapies are often restricted to a subset of patients.
- Predictive biomarkers: In some cases, mutated cancer genes are potent biomarkers for response or resistance to targeted agents.
- By linking drug activity to the functional complexity of cancer genomes, systematic pharmacogenomic profiling in cancer cell lines provides a powerful biomarker discovery platform to guide rational cancer therapeutic strategies.

## Pharmacogenomics



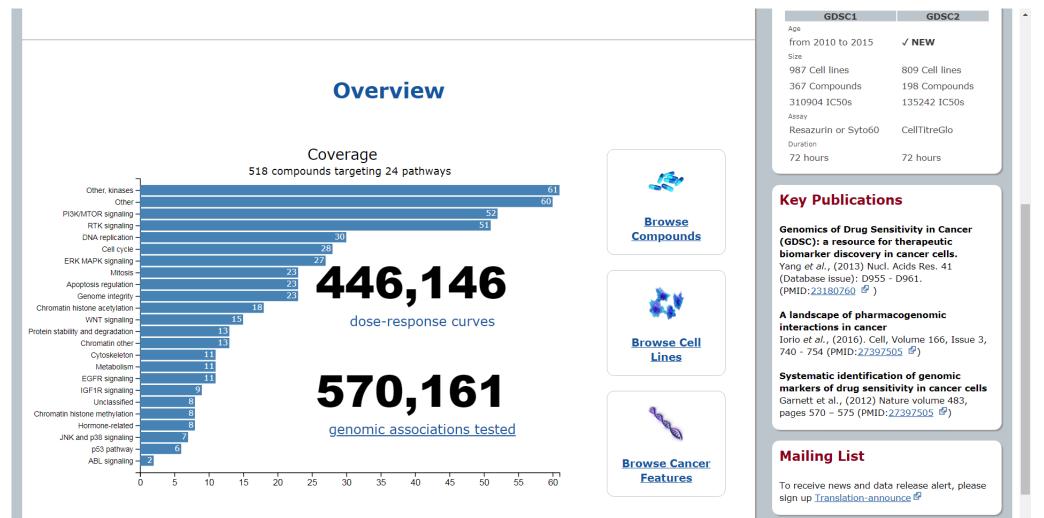
- Pharmacogenomics: the study of how genes affect a person's response to drugs.
- This relatively new field combines pharmacology (the science of drugs) and genomics (the study of genes and their functions) to develop **effective**, **safe** medications and **doses** that will be tailored to a person's genetic makeup.
- Pharmacogenetics: how variation in one single gene influences the response to a single drug.
- **Pharmacogenomics**: a broader term, which studies how all of the genes (the genome) can influence responses to drugs.

## The Genomics of Drug Sensitivity in Cancer

 The Genomics of Drug Sensitivity in Cancer Project is a collaboration between the Cancer Genome Project at the Wellcome Sanger Institute (UK) and the Center for Molecular Therapeutics, Massachusetts General Hospital Cancer Center (USA). This work is funded by Wellcome. **Primary tumors:** Cancer genomic alterations identified in **11,289 tumors** from 29 tissues (Somatic mutations, Copy number alterations, DNA methylation)

Cancer cells: 1,001 molecularly annotated human cancer cell lines

**Drugs:** 518 drugs were tested on cells



- Classic oncogene addiction paradigms were modified by additional tissue-specific or expression biomarkers.
- We find that cell lines faithfully recapitulate oncogenic alterations identified in tumors, find that
  many of these associate with drug sensitivity/resistance, and highlight the importance of
  tissue lineage in mediating drug response.
- We found that **mutated cancer genes were associated with cellular response** to most currently available cancer drugs.
- Our analysis and datasets are rich resources to link genotypes with cellular phenotypes and to identify therapeutic options for selected cancer sub-populations.

### Gene-drug interactions in cancer cells in vitro

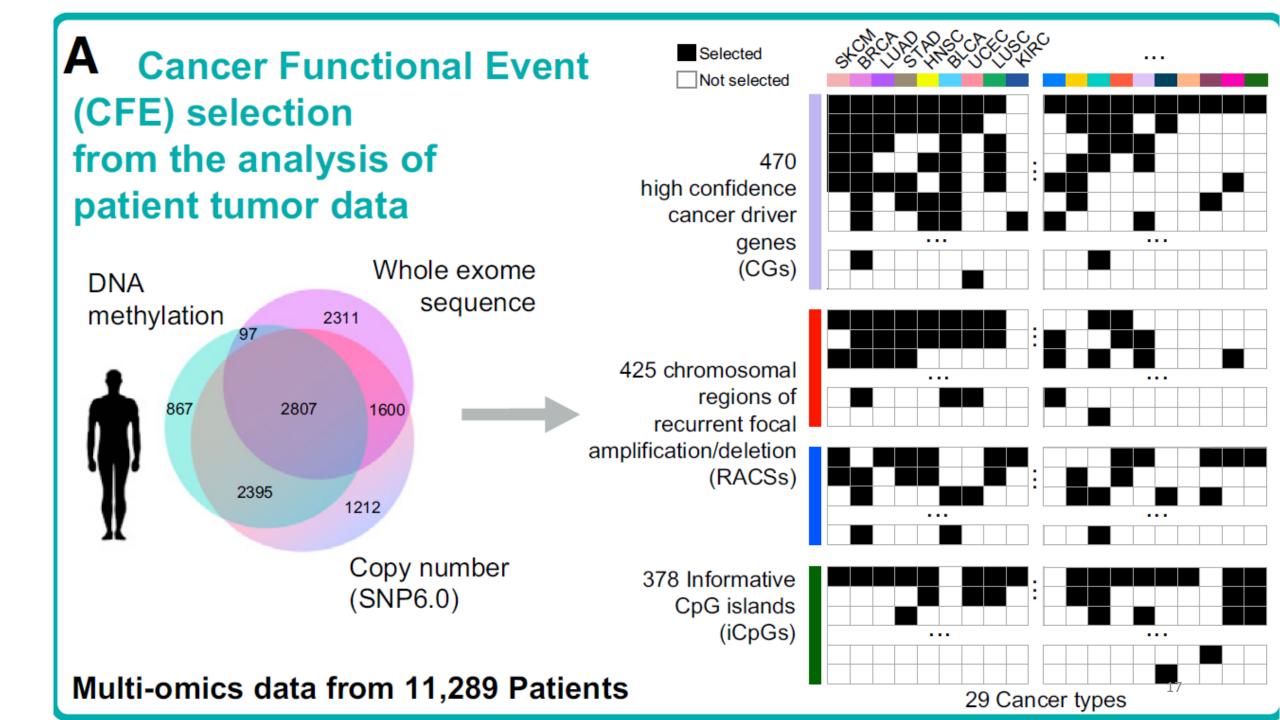
- Clinical trials are complex and expensive, and **pre-clinical data** that helps stratify patients can dramatically increase the likelihood of success during clinical development (Cook et al., 2014; Nelson et al., 2015).
- Thus, pre-clinical biological models that, as much as reasonably possible, capture both the molecular features of cancer and the diversity of therapeutic responses are a necessity.
- Human cancer cell lines are a facile experimental model and are widely used for drug development.
- Large-scale drug sensitivity screens in cancer cell lines have been used to identify clinically meaningful gene-drug interactions.
- In the **past**: imperfect understanding of the landscape of cancer driver genes
- Now: view drug sensitivity through the lens of clinically relevant oncogenic alterations

### GDSC GOALS

- (1) the extent to which cancer cell lines recapitulate oncogenic alterations in primary tumors
- (2) which oncogenic alterations associate with drug sensitivity
- (3) whether logic **combinations** of multiple alterations better explain drug sensitivity
- (4) the relative contribution of **different molecular data types**, either individually or in combination, in **predicting drug response**.

# "Cancer functional events" (CFEs)

- (1) Cancer genes (CGs) for which the mutation pattern in wholeexome sequencing (WES) data is consistent with positive selection
- (2) focal recurrently aberrant **copy number** segments (RACSs) from SNP6 array profiles
- 3) **Hypermethylated** informative 5'C-phosphate-G-3' sites in gene promoters (iCpGs) from DNA methylation data



- This identified 461 unique pan-cancer genes.
- We mined the COSMIC database to identify likely driver mutations
- Most tumors harbored only a few driver mutations (median n = 2, range 0-64), consistent with previous reports (Kandoth et al., 2013; Vogelstein et al., 2013).

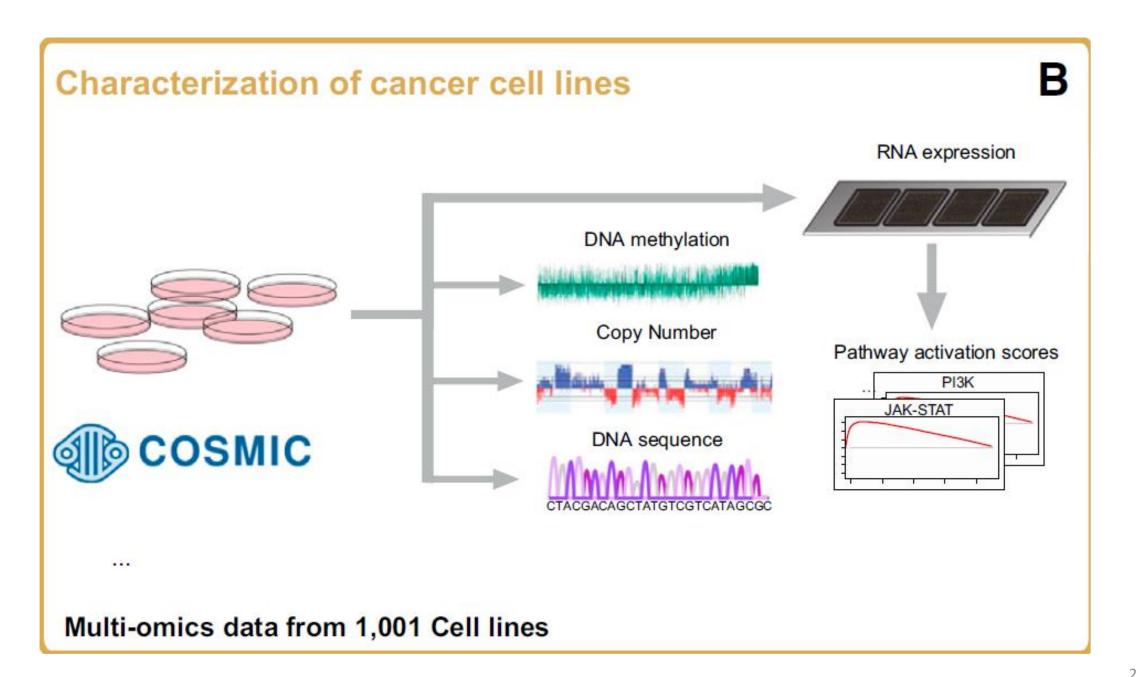
## Recurrently aberrant copy number segments (RACSs)

- 851 cancer-specific RACSs were gained (286 segments) or lost (565 segments), with a median of 19 RACSs per tumor type.
- The median number of genes within each RACS was 15 for amplified regions and one for deleted regions.
- The majority of known driver gene amplifications (e.g., EGFR, ERBB2, MET, and MYC) and homozygous deletions (e.g., CDKN2A, PTEN, and RB1) were captured, with 320 RACSs (38%) containing at least one known putative cancer driver gene, in addition to 531 RACSs (62%) without known driver genes.

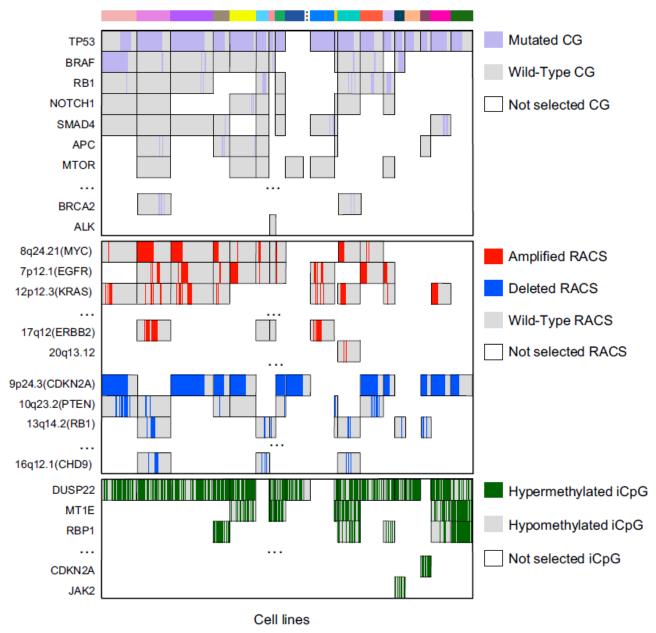
- iCpGs were identified using DNA methylation array data for 6,166 tumor samples spanning 21 cancer types.
- We defined 378 iCpGs based on a multimodal distribution of their methylation signal in at least one cancer type (Tables S2H and S2I).
- This also established a discretization threshold used to define such regions as hyper-methylated in the cell lines
- In total, our multidimensional analysis of >11,000 patient tumor samples identified 1,699 cancer-specific CFEs, which were further merged into 1,273 unique pan-cancer CFEs.

## Oncogenic Alterations in Patient Tumors Are Conserved across Cell Lines

- ▶ Next, we assessed the extent to which the mutational landscape of cancer cell lines captures that seen in primary tumors.
- We utilized a panel of 1,001 human cancer cell lines analyzed through WES (n = 1,001), copy number (n = 996), gene expression (n = 968), and DNA methylation (n = 957) (<a href="https://cancer.sanger.ac.uk/cell lines/cbrowse/all">https://cancer.sanger.ac.uk/cell lines/cbrowse/all</a>) and which we reclassified according to the TCGA tissue labels.
- Molecular alterations identified in cell lines were **filtered using the CFEs identified in the primary tumor samples**, providing a set of clinically relevant CFEs for the cell lines (Figure 1C).

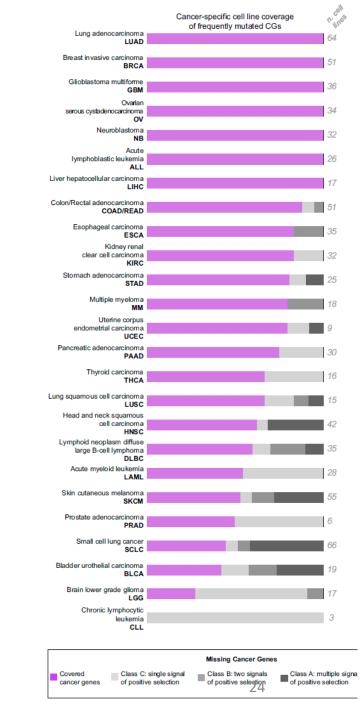


#### C Clinically relevant CFEs in cancer cell lines



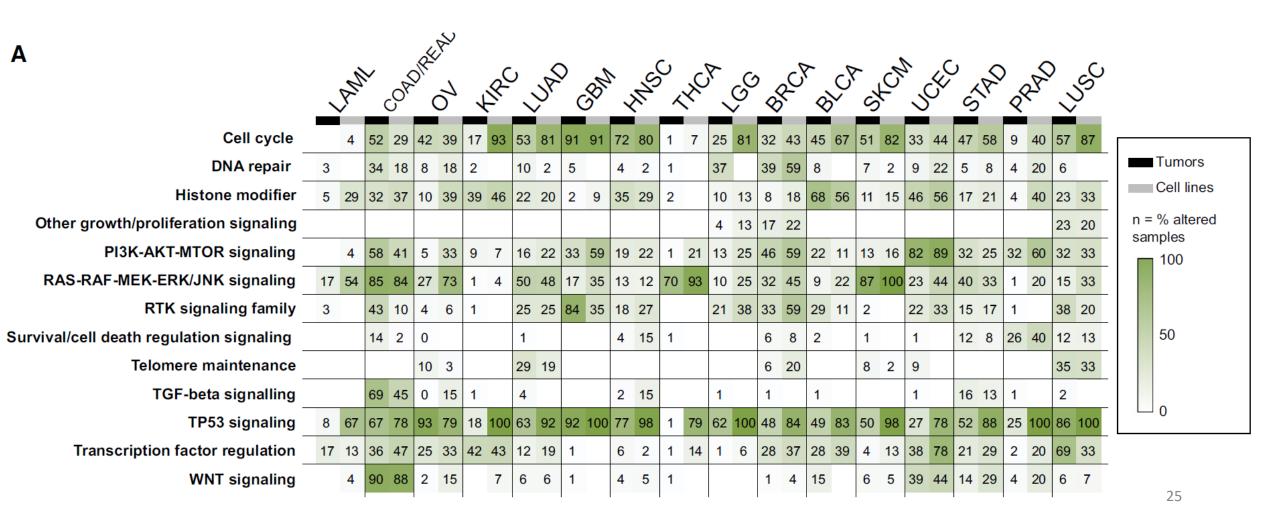
### CFEs in cell lines

- Of the 1,273 pan-cancer CFEs identified in patient tumors, 1,063 (84%) occurred in at least one cell line, and 1,002 (79%) occurred in at least three (Figure 2A).
- This concordance was greatest for the RACSs (100% of 425), followed by iCpGs (338 of 378, 89%) and CGs (300 of 470).
- When considering cancer-specific CFEs, concordance was highest for CFEs occurring in at least 5% of patients (median of 86% of CFEs covered across cancer types).
- Coverage of CFEs varied by cancer type.



 Alterations in 13 canonical cancer-associated pathways was highly correlated between cell lines and tumors of the same cancer type.

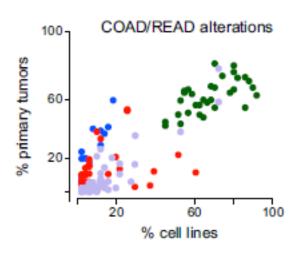
(See Excel file)



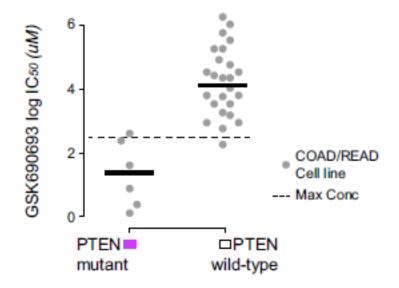
## 4D

#### Analyses

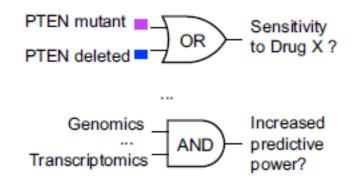
1. Comparison of cell lines and primary tumors across cancer types



2. Identification of clinically relevant markers of drug response



3. Predictive ability assessment of individual CFE/data-type and their combinations



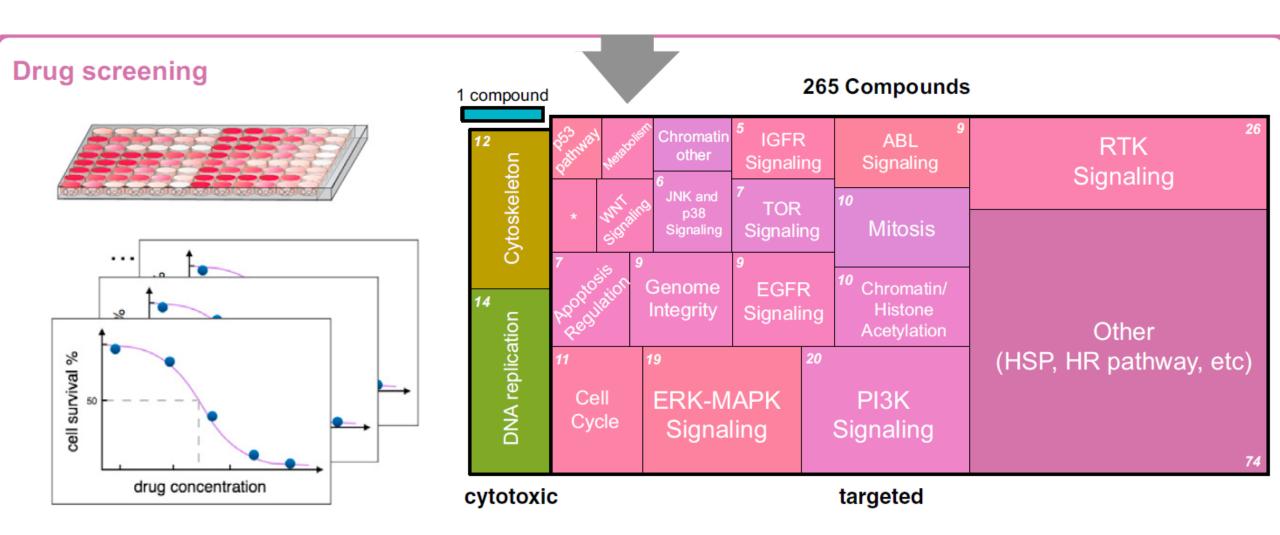
http://www.cancerrxgene.org



 These results show that a sufficiently large panel of cell lines is able to capture individual clinically relevant genomic alterations, in addition to pathway alterations and global signatures of driver events.

## A Therapeutic Landscape of Human Cancers Modeling Pharmacogenomic Interactions

- To investigate how CFEs detected in primary tumors impact drug response.
- Cell lines underwent extensive drug sensitivity profiling.
- The effect of each drug on cell number was used to model sensitivity as IC50 (drug concentration that reduces viability by 50%) or AUC (area under the dose-response curve) values (Tables S4A and S4B).



212,774 Compound/Cell line dose-response curves

<sup>\*</sup> Chromatin/Histone Methylation

 Screened compounds included cytotoxics (n = 19) and targeted agents (n = 242) selected against 20 key pathways and cellular processes in cancer biology

• These 265 compounds include clinical drugs (n = 48), drugs currently in clinical development (n = 76), and experimental compounds (n = 141).

## Cell viability assays

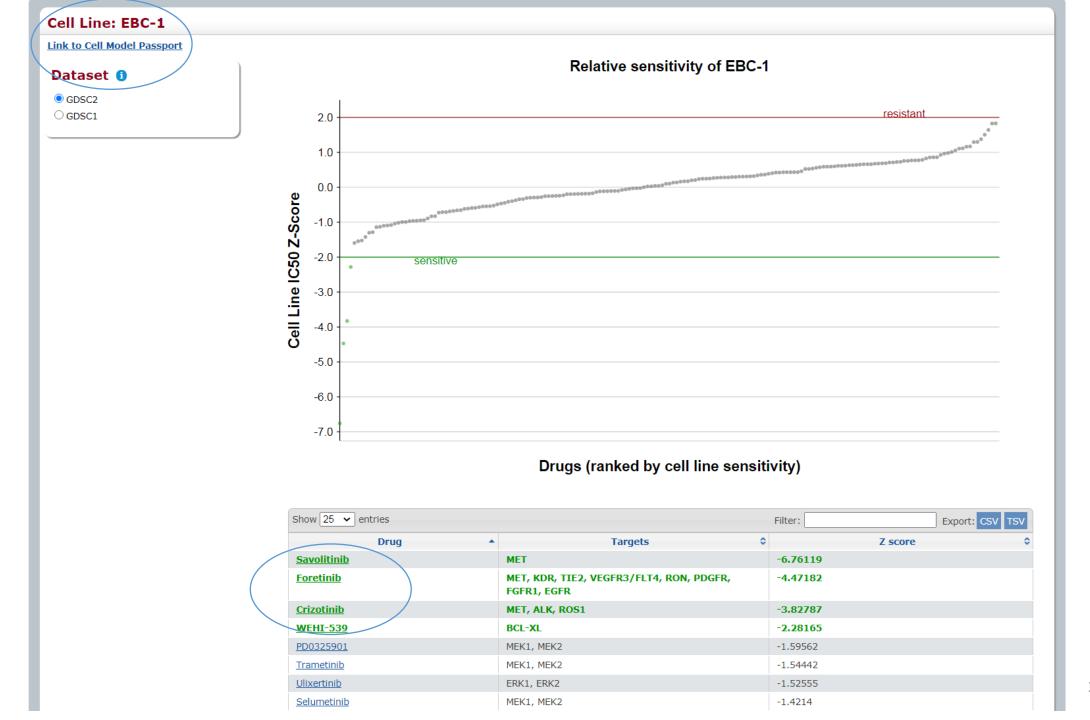
- Cells were seeded in either 96-well or 384-well microplates in medium supplemented with 5% FBS and penicillin/streptavidin.
- The optimal cell number for each cell line was determined to ensure that each was in growth phase at the end of the assay (70% confluency).
- Adherent cell lines were plated 1 day before treatment with a **9-point twofold dilution** series of each compound using liquid handling robotics, and assayed at a **72-h time point**.
- Cells were fixed in 4% formaldehyde for 30 min and then stained with 1 mM of the fluorescent nucleic acid stain Syto60 (Invitrogen) for 1 h.
- Suspension cell lines were treated with compound immediately following plating, incubated for 72 h, and then stained with 55 mgml
   resazurin (Sigma) prepared in glutathione-free media for 4 h.
- Quantification of fluorescent signal intensity was performed using a fluorescent plate reader at excitation and emission wavelengths of
   630/695nM for Syto60, and 535/595nM for resazurin.
- All screening plates were subjected to stringent quality control measures and a Z-factor score comparing negative and positive control wells was calculated.

### **Datasets**

GDSC1	GDSC2
Age	
from 2010 to 2015	√ NEW
Size	
987 Cell lines	809 Cell lines
367 Compounds	198
	Compounds
310904 IC50s	135242 IC50s
Assay	
Resazurin or	CellTitreGlo
Syto60	
Duration	
72 hours	72 hours

 Cluster analysis based on AUC values confirmed that compounds with overlapping nominal targets or targeting the same process/pathway had similar activity profiles (See Excel file, compound clustering)

- We used three distinct analytical frameworks to define the contribution of CFEs to the prediction of drug sensitivity
- ANOVA was used to identify single CFEs as markers of drug response.
- Logic models identified combinations of CFEs that improve the prediction of drug response.
- We used machine-learning algorithms to assess the contribution of each molecular data type (CGs, RACS, iCpGs, and gene expression) in explaining variation in drug response

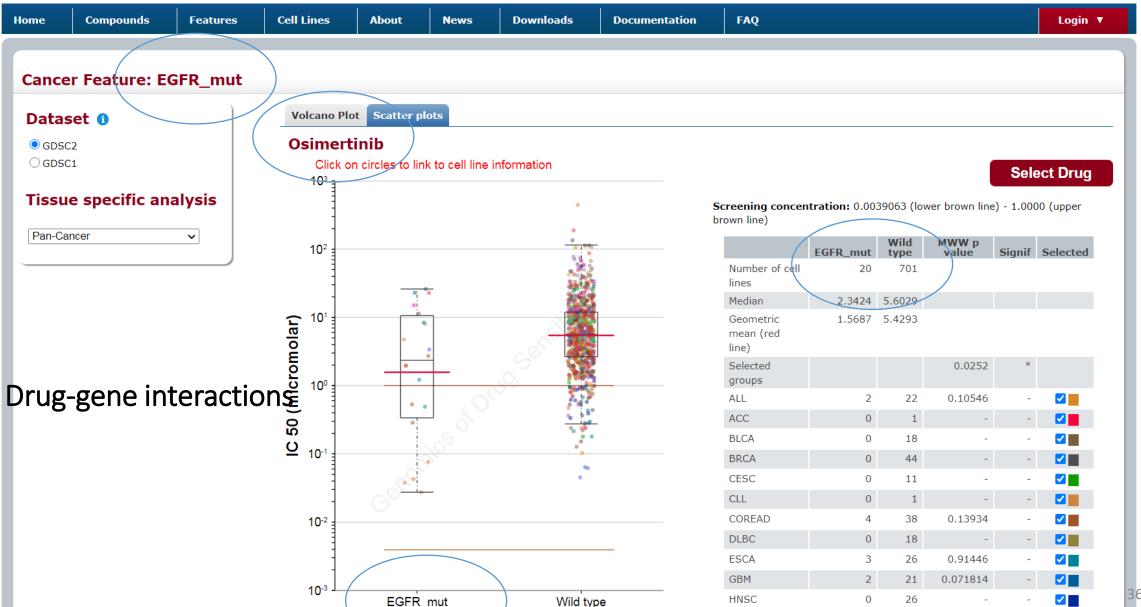


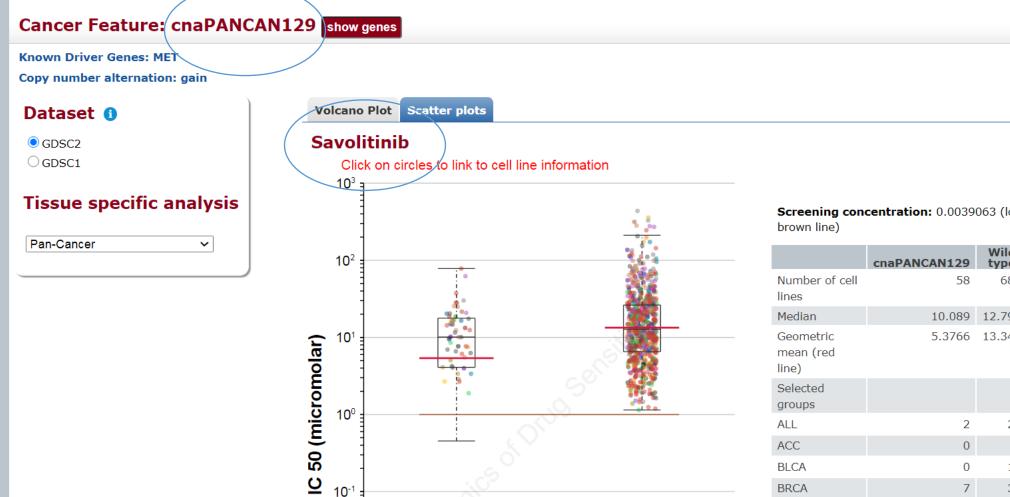


#### **Genomics of Drug Sensitivity in Cancer**









10<sup>-2</sup>

10<sup>-3</sup>

cnaPANCAN129

Wild type

**Select Drug** 

Screening concentration: 0.0039063 (lower brown line) - 1.0000 (upper

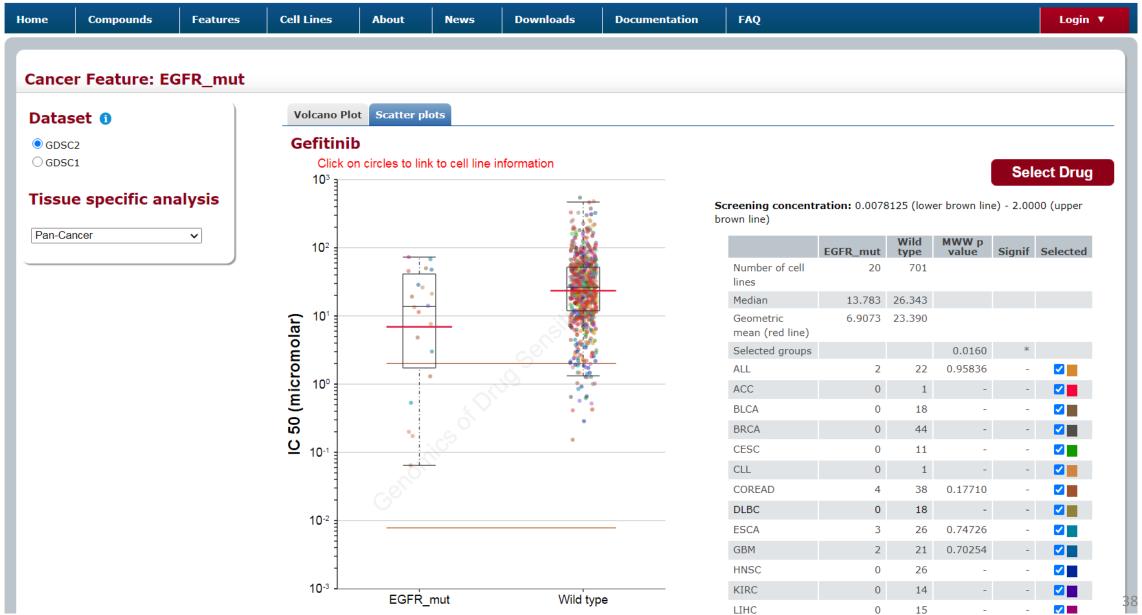
	cnaPANCAN129	Wild type	MWW p value	Signif	Selected
Number of cell lines	58	688			
Median	10.089	12.793			
Geometric mean (red line)	5.3766	13.341			
Selected groups			0.0107	*	
ALL	2	21	0.62344	-	<b>✓</b>
ACC	0	1	-	-	
BLCA	0	18	-	-	<b>✓</b>
BRCA	7	37	0.10174	-	<b>✓</b>
CESC	1	12	0.76923	-	<b>✓</b>
CLL	0	2	-	-	
COREAD	3	39	0.84512	-	<b>✓</b>
DLBC	1	18	0.73684	-	
ESCA	2	27	0.022562	*	<b>✓</b>
GBM	2	22	0.27278	-	<b>✓</b>
HNSC	1	25	0.35064	-	<b>✓</b>
KIRC	2	14	0.066667	-	<b>2</b>
LIHC	1	12	0.92308	-	37



#### **Genomics of Drug Sensitivity in Cancer**



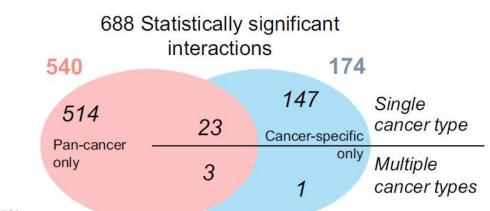


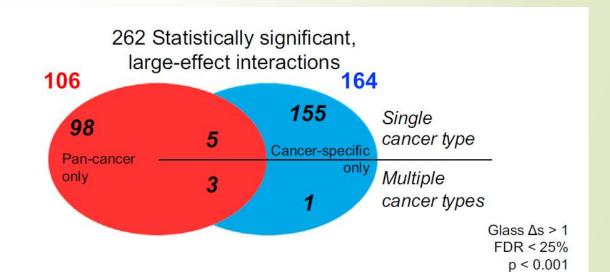


#### 39

## ANOVA Analysis Defines a Landscape of Pharmacogenomic Interactions

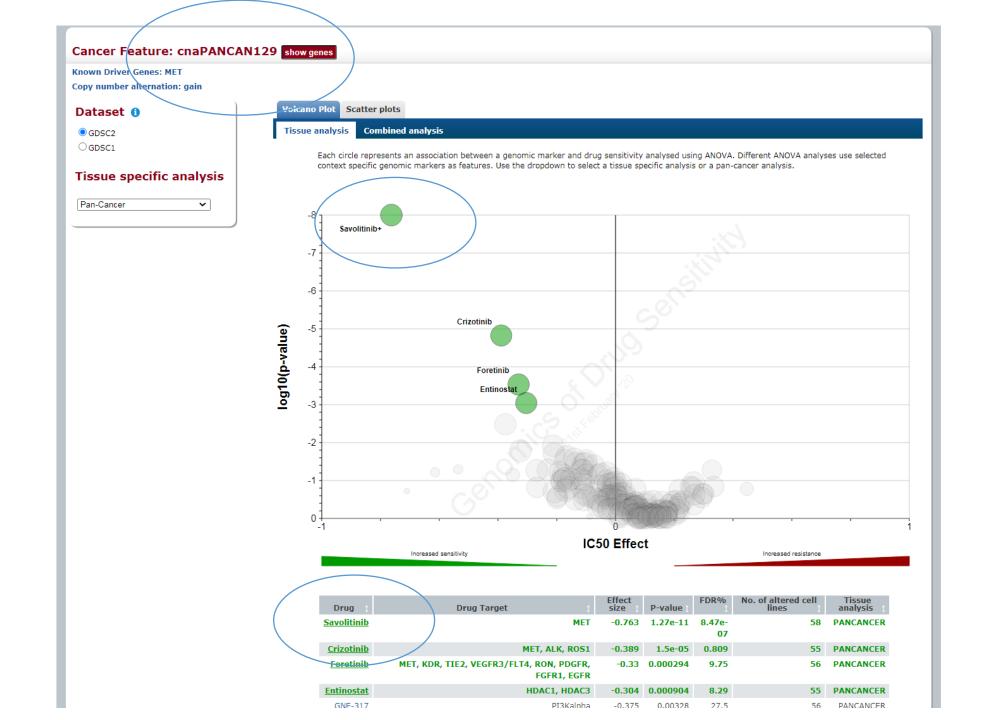
- For pan-cancer ANOVA, the set of CFEs included 267 CGs, 407 RACSs, and three gene fusions (BCR-ABL, EWSR1-FLI1, and EWSR1-X).
- Overall, for the 265 compounds, we identified 688 statistically significant interactions between unique CFE-drug pairs (p value < 103 at a false discovery rate [FDR] < 25%; Figure 4A), with 540 pan-cancer and 174 cancer-specific hits.
- A subset of 262 CFE-drug pairs was additionally defined as large-effect interactions.





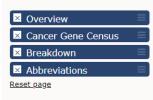
FDR < 25% p < 0.001

A



GRCh38 · COSMIC v92

#### Census



#### Overview

The Cancer Gene Census (CGC) is an ongoing effort to catalogue those genes which contain mutations that have been causally implicated in cancer and explain how dysfunction of these genes drives cancer. The content, the structure, and the curation process of the Cancer Gene Census was described and published in Nature Reviews Cancer &.

The census is not static, instead it is updated when new evidence comes to light. In particular we are grateful to Felix Mitelman and his colleagues in providing information on more genes involved in uncommon translocations in leukaemias and lymphomas. Currently, more than 1% of all human genes are implicated via mutation in cancer. Of these, approximately 90% contain somatic mutations in cancer, 20% bear germline mutations that predispose an individual to cancer and 10% show both somatic and germline mutations.

#### Census tiers

Genes in the Cancer Gene Census are divided into two groups, or tiers.

#### Tier 1

To be classified into Tier 1, a gene must possess a documented activity relevant to cancer, along with evidence of mutations in cancer which change the activity of the gene product in a way that promotes oncogenic transformation. We also consider the existence of somatic mutation patterns across cancer samples gathered in COSMIC. For instance, tumour suppressor genes often show a broad range of inactivating mutations and dominant oncogenes usually demonstrate well defined hotspots of missense mutations. Genes involved in oncogenic fusions are included in Tier 1 when changes to their function caused by the fusion drives oncogenic transformation, or in cases when they provide regulatory elements to their partners (e.g. active promoter or dimerisation domain).

#### Tier 2

A new section of the Census, which consists of genes with strong indications of a role in cancer but with less extensive available evidence. These are generally more recent targets, where the body of evidence supporting their role is still emerging.

#### Hallmarks

New overviews of cancer gene function focused on hallmarks of cancer pull together manually curated information on the function of proteins coded by cancer genes and summarise the data in simple graphical form. They present a condensed overview of most relevant facts with quick access to the literature source, and define whether a gene has a stimulating or suppressive effect via individual cancer hallmarks. Genes with the hallmark descriptions available are marked with the hallmark icon, that when clicked, opens the hallmark page. Hallmark descriptions will be expanded to encompass more genes and updated on regular basis.

#### Cancer Gene Census



TCGA Label	Definition		
ACC	Adrenocortical carcinoma		
ALL	Acute lymphoblastic leukemia		
BLCA	Bladder Urothelial Carcinoma		
BRCA	Breast invasive carcinoma		
CESC	Cervical squamous cell carcinoma and endocervical adenocarcinoma		
CLL	Chronic Lymphocytic Leukemia		
COREAD	Colon adenocarcinoma and Rectum adenocarcinoma		
DLBC	Lymphoid Neoplasm Diffuse Large B-cell Lymphoma		
ESCA	Esophageal carcinoma		
GBM	Glioblastoma multiforme		
HNSC	Head and Neck squamous cell carcinoma		
KIRC	Kidney renal clear cell carcinoma		
LAML	Acute Myeloid Leukemia		
LCML	Chronic Myelogenous Leukemia		
LGG	Brain Lower Grade Glioma		
LIHC	Liver hepatocellular carcinoma		
LUAD	Lung adenocarcinoma		
LUSC	Lung squamous cell carcinoma		
MB	Medulloblastoma		
MESO	Mesothelioma		
MM	Multiple Myeloma		
NB	Neuroblastoma		
OV	Ovarian serous cystadenocarcinoma		
PAAD	Pancreatic adenocarcinoma		
PRAD	Prostate adenocarcinoma		
SCLC	Small Cell Lung Cancer		
SKCM	Skin Cutaneous Melanoma		
STAD	Stomach adenocarcinoma		
THCA	Thyroid carcinoma		
UCEC	Uterine Corpus Endometrial Carcinoma		

#### Drug-gene interactions



Dataset (1)

OGDSC2

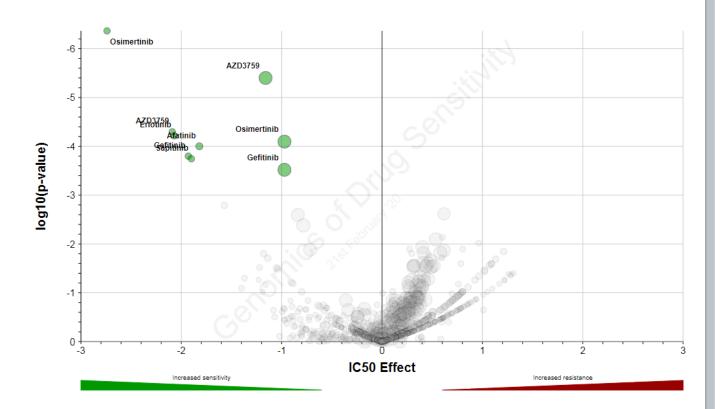
○GDSC1

Volcano Plot Scatter plots

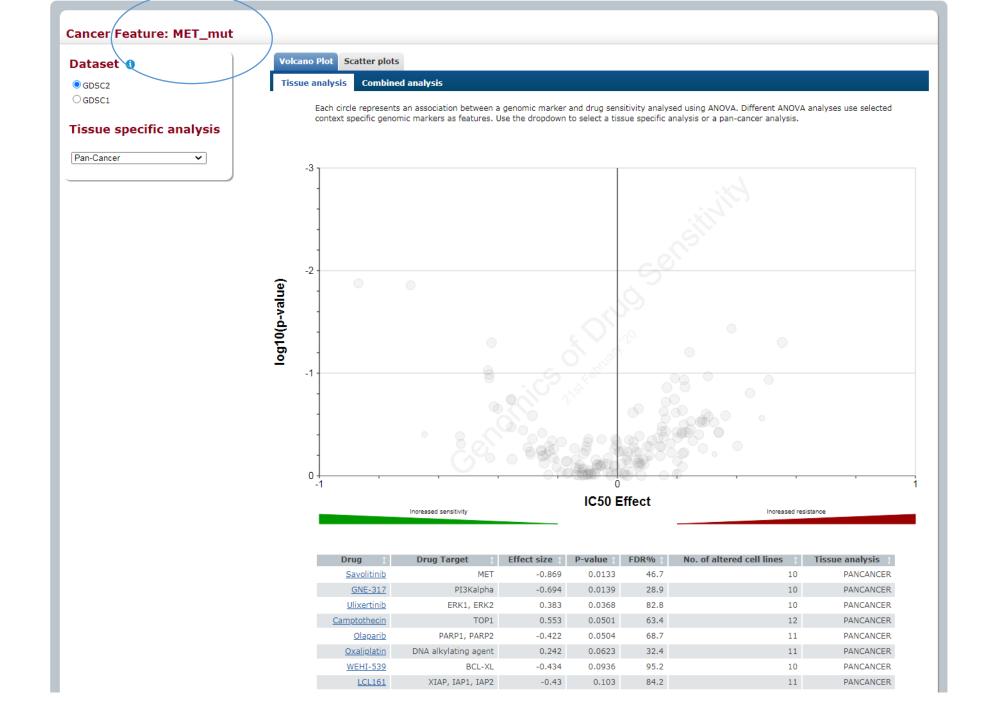
Tissue analysis

**Combined analysis** 

Associations from all tissue specific and pan-cancer ANOVA analyses overlaid. Hover the pointer over each circle to reveal the context-specific details of an association.



Drug	Drug Target	Effect size	P-value 1	FDR% ‡	No. of altered cell lines	Tissue analysis
<u>Osimertinib</u>	EGFR	-2.74	4.32e-07	0.00129	5	LUAD
AZD3759	EGFR	-1.16	3.98e-06	0.264	20	PANCANCER
AZD3759	EGFR	-2.09	5.04e-05	0.101	5	LUAD
<u>Erlotinib</u>	EGFR	-2.07	6.07e-05	0.207	5	LUAD
<u>Osimertinib</u>	EGFR	-0.972	8.03e-05	1.78	20	PANCANCER
<u>Afatinib</u>	ERBB2, EGFR	-1.82	0.0001	0	6	LUAD
<u>Gefitinib</u>	EGFR	-1.93	0.000159	0.54	5	LUAD
<u>Sapitinib</u>	EGFR, ERBB2, ERBB3	-1.9	0.00018	0.647	5	43 LUAD
<u>Gefitinib</u>	EGFR	-0.972	0.000303	6.05	20	PANCANCER





#### **Genomics of Drug Sensitivity in Cancer**







You are here: Home > Browse Models > LNCaP-Clone-FGC

Model Information

#### LNCaP-Clone-FGC

#### Overview

Name(s) LNCaP-Clone-FGC (LNCaP-FGC, LNCaP-ATCC)

Tissue Prostate

Cancer Type Prostate Carcinoma

Tissue Status Metastasis

Sample Site Left supraclavicular lymph node

Cancer Type Details Prostate Carcinoma (NCIT C4863)

#### Clinical Information

#### Gender Male

Ethnicity White

Smoker Status Unknown

Age at sampling 50

Prior treatment Unknown

Prior treatment details

#### Derivation

Model Type Cell Line

Growth Properties Semi-Adherent

Model treatment

Year Generated 1977

#### Model Genomics

MSI Status MSI Ploidy 3.776

Mutations per Mb 90.833

#### Dataset Availability

Raw Whole Exome Sequencing (BAM / CRAM) - Source: Sanger

Processed LNCaP-Clone-FGC.cave.annot.vcf.gz (1.25 MB) - Source: Sanger

Processed LNCaP-Clone-FGC.pindel.annot.vcf.gz (1.53 MB) - source: Sanger

#### RNASeq

Processed rnaseq\_20191101.zip (58.73 MB)

#### Microarray Gene Expression

♠ Whole Exome Sequencing

Raw Microarray Gene Expression (CEL) - Source: Sanger

#### Copy Number Variation

Processed cnv\_20191101.zip (15.85 MB)

Raw Copy Number Variation (BAM / CRAM) - Source: Sanger

#### PDNA Methylation

Raw DNA Methylation (IDAT) - Source: Sanger

#### ■ Fusion data

Processed fusions\_latest.csv.gz (185.56 kB) - Source: Sanger

#### Drug Sensitivity Data

Link to Genomics of Drug Sensitivity in Cancer

#### Cancer Driver Mutations

Note: Displays all mutated cancer driver genes for this model. Font size indicates the frequency of the mutation within the model's tissue type. See cancer driver list documentation for annotation details.

IC50 (µM)

Crispr KO - highlights from Project SCORE

#### View LNCaP-Clone-FGC on the Project SCORE website

Top-5 genes with highest model-specific effect size\*

Gene Effect size p-value (Link to Project SCORE) (fold change) (MAGeCK FDR) Drug Sensitivity - highlights from CancerRxGene.org

#### View drug sensitivity for LNCaP-Clone-FGC on CancerRxGene.org

Top-5 drugs with lowest z-score

**Drug Name Drug Targets** (Link to GDSC)

Z Score

Dataset

https://score.depmap.sanger.ac.uk/

https://depmap.sanger.ac.uk/

• <a href="https://depmap.sanger.ac.uk/programmes/">https://depmap.sanger.ac.uk/programmes/</a> (Drug response)